

Pentamidine Inhibits Ovarian Cancer Cell Proliferation and Migration by Maintaining Stability of PTEN in vitro

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Purpose: Pentamidine is an anti-protozoal cationic aromatic diamidine drug and has been reported to exhibit anticancer properties. We aimed to identify the effect of pentamidine on proliferation and migration of human ovarian cancer (OC) cell lines and the related mechanisms.

Methods: HO8910 and Caov3 ovarian cancer cells were treated with pentamidine. MTS and colony formation assays were used to detect the proliferation ability of cells. The migration of cells was detected using wound healing and transwell assays. The protein levels of PTEN, phosphorylated Akt, Akt, N-cadherin, E-cadherin and snail were detected by Western blotting. Immunoprecipitation and Western blotting were used to detect ubiquitination levels of PTEN.

Results: Our findings revealed that pentamidine inhibited both proliferation and migration of OC cells. Further investigation found that pentamidine increased the protein expression of PTEN and reduced phosphorylation levels of AKT in OC cells. Pentamidine treatment modulated PTEN stability through the ubiquitin/proteasome pathway. In addition, pentamidine inhibited the expression of N-cadherin and snail, and increased E-cadherin expression in a dose-dependent manner.

Conclusion: Pentamidine is involved in the maintenance of PTEN protein stability and suppresses proliferation and migration of OC cells.

Keywords: pentamidine, ovarian cancer, PTEN

Introduction

As of 2018, ovarian cancer (OC) was the seventh most common cancer worldwide in women, with around 240,000 new cases.¹ Ovarian cancer is the second most common malignancy after breast cancer in women over the age of 40, particularly in developed countries.¹ At present, the clinical treatment options for OC mainly include surgery, radiation therapy, chemotherapy and targeted therapy or their combination. Although there have been many innovations in the treatment of ovarian cancer in the past decades, five-year relative survival is below 45% and, unlike other common cancer types, the proportion of women who die from their disease has not improved substantially over time.² Thus, exploring novel molecular therapy and new drug is necessary and urgent.

PTEN (phosphatase and tensin homologue deleted on chromosome ten), also known as TEP1 (TGF β -regulated and epithelial cell-enriched phosphatase) or MMAC1 (mutated in multiple advanced cancers), is a well-known tumor suppressor

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gene.³ PTEN is a dual phosphatase with activity on lipids and proteins which can dephosphorylate phosphatidylinositol (3,4,5)-trisphosphate (PIP3) to phosphatidylinositol (4,5)-trisphosphate (PIP2) and antagonize the phosphatidylinositol 3-kinase (PI3K)/Akt oncogenic pathway, thereby inhibiting cell proliferation, survival and migration.^{3,4} Recent studies showed that chemotherapy drugs, such as cisplatin, doxorubicin and paclitaxel, could induce tumor cells apoptosis through enhanced PTEN activity and overexpression of PTEN is able to enhance the anti-tumor effect of chemotherapy drugs in many malignant tumors including OC.⁵ PTEN expression was precisely regulated on its post-transcriptional modifications, including ubiquitylation. Loss or mutation of PTEN has frequently been found in ovarian cancer and is associated with the worse prognosis of cancer patients.⁶ Therefore, pharmacological control of PTEN expression and activity might be an important strategy for cancer treatment.

Pentamidine is a positively charged aromatic diamine that has a long history of its involvement in the clinical treatment of African trypanosomiasis, antimonial-resistant leishmaniasis and babesiosis as well as the prophylaxis of pneumocystis carinii pneumonia in acquired immune deficiency syndrome (AIDS) patients for several decades.^{7–11} Recently, several reports suggested that pentamidine has antitumor potential. In fact, pentamidine has been shown to have anti-proliferative effects on various human cancer cell types such as melanoma, prostate, ovarian, colon, breast, lung, and cervical cancers *in vitro*.^{12–15} However, anticancer targets of pentamidine and molecular mechanism have not been fully defined in OC.

In this study, our findings demonstrated that pentamidine suppresses cell proliferation and migration in ovarian cells. Moreover, we provided evidence to show pentamidine modulates PTEN stability by ubiquitin/proteasome pathway in OC cells. Furthermore, pentamidine treatment upregulates epithelial markers and downregulated mesenchymal markers in OC cells. Therefore, increasing PTEN stability with pentamidine may serve as its anticancer underlying molecular mechanism in OC.

Materials and Methods

Chemicals and Antibodies

Pentamidine was obtained from MedChemExpress (Shanghai China). Cycloheximide (CHX) was obtained from Sigma-Aldrich (St. Louis, MO, USA). MG132 was

obtained from Cell Signaling Technology (Beverly, MA, USA).

Antibodies against Phospho-AKT (Ser473, Cat. #. 4060T, 1:1000), AKT (Cat. #. 4691T, 1:1000), E-Cadherin (Cat. #. 3195T, 1:1000) and snail (Cat. #. 3879P, 1:1000) were obtained from Cell Signaling Technology. Antibodies against PTEN (Cat. #. 22034-1-AP, 1:2000), N-Cadherin (Cat. #. 62219-1-Ig, 1:2000), GAPDH (Cat. #. 60004-1-Ig, 1:20,000) and ubiquitin (Cat. #. 10201-2-AP, 1:500) were obtained from Proteintech.

Cell Lines and Cell Culture

The human ovarian cancer cell lines HO8910 and Caov3 were obtained from American Type Culture Collection (ATCC). HO8910 cells were cultured in RPMI1640 (GIBCO-BRL) and Caov3 cells were cultured in Dulbecco's modified Eagle's medium (DMEM). The culture medium was supplemented with 10% fetal bovine serum (FBS), 100 units/mL streptomycin and penicillin at 37°C under 5% (v/v) CO₂.

Cell Proliferation Assay

Cells were seeded into 96-well plates at a density of 3×10^3 cells/well and cultured overnight. Then, cells were treated with different concentrations of pentamidine for 24 h. Finally, growth viability of cells was assayed using the one solution cell proliferation assay (MTS assay) (Promega).

Colony Formation Assay

The cells were seeded in 6-well plates at the density of 1000 cells/well and incubated overnight. After culturing the cells with different concentrations of pentamidine for 7 days, the colonies were fixed with 4% paraformaldehyde and stained with crystal violet. The numbers of cell colonies were counted using the ImageJ software.

Wound Healing Assay

Cells were seeded into 6-well culture plates. When the cells reached confluent, cells were washed with phosphate buffer saline (PBS) and cultured with or without pentamidine treatment in RPMI1640 medium or DMEM medium without serum to minimize cell proliferation. Then, a sterile plastic pipette tip was used to make a straight scratch. After scratching, the well was gently washed twice with PBS to remove detached cells. The scratch was examined and photographed under a light microscope at 0, 24 and 48 hours.

Transwell Assay

Cell migration assays were performed using 24-well cell culture plates with 8- μ m pore filter inserts (Corning). About 3×10^4 cells in 100 μ L serum-free medium were placed in upper chamber, and 600 μ L RPMI1640 medium supplemented with 10% FBS was added to the lower chamber. Pentamidine or vehicle was added into both upper chamber and lower chamber. After 6 hours incubation at 37°C and 5% CO₂, cells on the upper surface were wiped away gently, whereas cells that migrated to the lower surface were fixed with 95% ethanol for 30 min and stained with 0.1% crystal violet for 30 min at room temperature. The cells were photographed and the migration cell numbers were counted in five random fields using ImageJ software (procedures: 1. open the target images with ImageJ 2. convert images to black and white 3. adjust threshold 4. separate the cells that are linked together with watershed 5. analyze particles 6. display results). The data are presented as mean $\pm$ standard deviation.

Real-Time Quantitative PCR

Total RNA was isolated from cells pre-treated with 20 μ mol/L pentamidine or vehicle for 24 hours using Trizol reagent (Invitrogen) and then reverse transcribed to cDNA using the PrimeScriptTM RT-PCR Kit (TaKaRa) according to the manufacturer's instructions. Real-time quantitative polymerase chain reaction (qPCR) was performed using the SYBR Premix Ex Taq kit (TaKaRa) on a Light Cycler 96 instrument (Roche Life Science). Gene expression levels were calculated relative to ribosome 18S rRNA. The sequences of the primers were as follows: PTEN (FOR): 5'-GCA GAA AGA CTT GAA GGC GTA-3', PTEN (REV): 5'-TTG GCG GTG TCA TAA TGT CT-3', 18S (FOR): 5'-TTG ACG GAA GGG CAC CAC CAG-3', 18S (REV): 5'-GCA CCA CCA CCC ACG GAA TCG-3'.

Western Blot and Immunoprecipitation

Cells with indicated treatments were lysed using RIPA buffer with protease cocktail inhibitor (APExBIO). The lysates were incubated on ice for 30 min with intermittent vortexing every 5 min, followed by centrifugation at 12,000g for 20 min at 4°C. Protein concentration was determined with BCA protein assay kit (Pierce). For Western blot, equivalent amounts of protein (10 μ g) from each sample were separated using 8% or 12% SDS-PAGE and transferred onto a polyvinylidene difluoride (PVDF)

membranes, then blocked with 5% skimmed milk. Membranes were incubated with primary antibodies overnight at 4°C. Subsequently, membranes were incubated with secondary antibodies for 1h by gentle shaking. Antibodies were dissolved in milk. Antibody binding was visualized using ECL chemiluminescence reagent (Thermo, MA, USA). For immunoprecipitation (IP), the whole cell lysates were extracted and clarified by centrifuging at 12,000 g for 20 min at 4 °C. The cell lysates were then incubated with indicated antibody for 2 h with gentle rotation followed by 30 μ L protein G-Sepharose overnight at 4°C. Then antibody-protein G-Sepharose-bound protein complexes were washed three times with TNE buffer. The bound immune complexes were analyzed by Western blotting.

Statistical Analysis

The data of all experiments were from three independent experiments and presented as mean $\pm$ SD. Statistical difference was determined by Student's *t* test between two sets of data. One-way analysis of variance (ANOVA) was applied to monitor the differences among groups. Tukey's test was used for multiple comparisons after ANOVA. The GraphPad 5.0 software was used and a difference of *P*<0.05 was considered statistically significant.

Results

Pentamidine Represses Growth and Proliferation of OC Cells

To determine whether pentamidine has any influence on ovarian cancer cell growth and proliferation, we first examined the cell viability upon pentamidine treatment by MTS assays and found that HO8910 and Caov3 cells were significantly inhibited in a dose-dependent manner, and this suppressive effect reached a peak at the dose of 20 μ mol/L (Figure 1A and B). Consistently, the results from cell colony formation experiments showed that the number of cell clones by HO8910 cells or Caov3 cells was significantly reduced in a dose-dependent manner after treatment with pentamidine for 7 days. In HO8910 cells, the number of clones decreased by 62.7%, 94.8% and 96.6%, respectively; in Caov3 cells, the number of clones decreased by 41.3%, 89.9% and 95.1%, respectively (Figure 1C–E). Taken together, these results suggested that pentamidine had a significant effect on reducing cell growth and proliferation of OC cells.

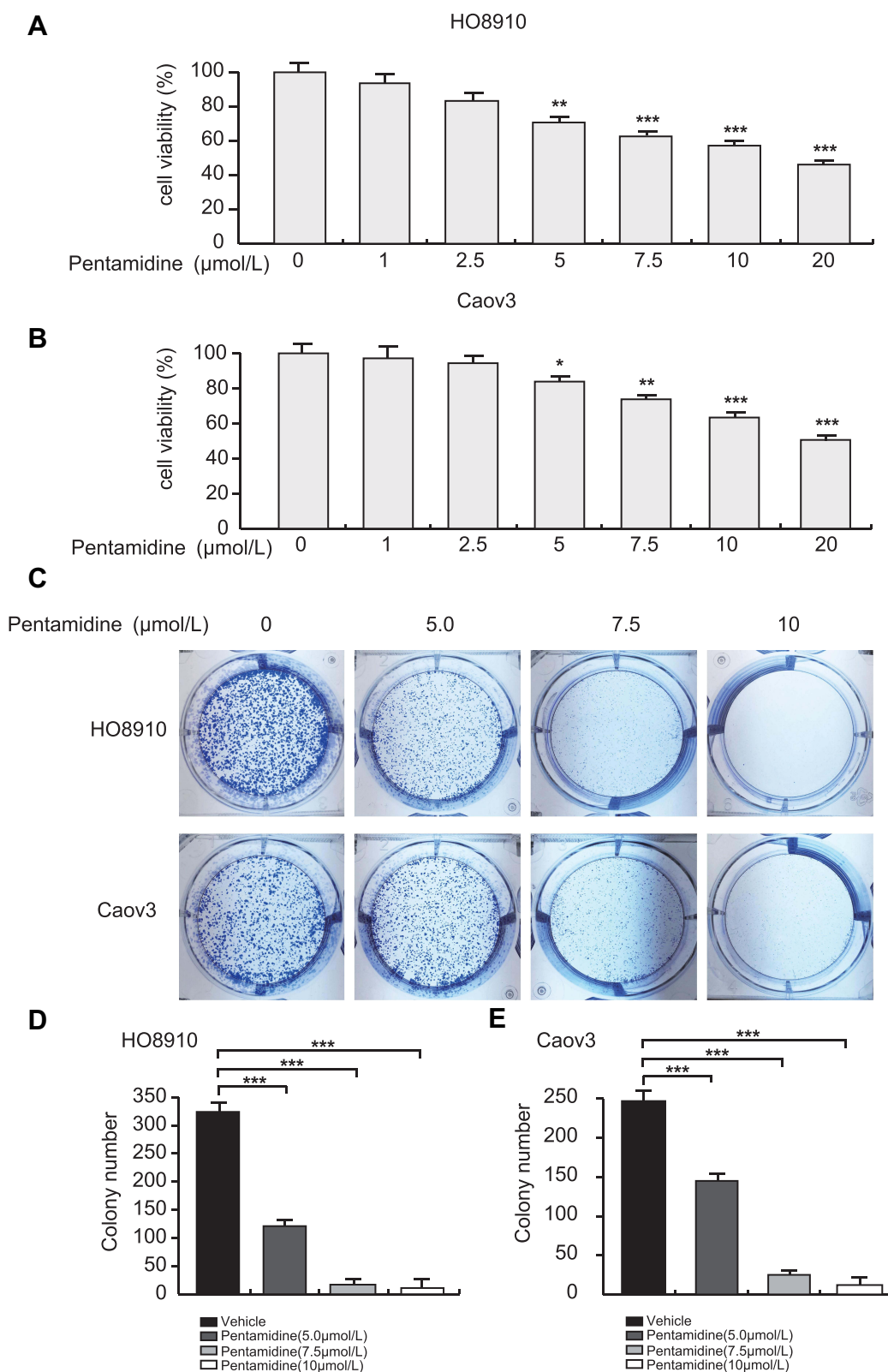


Figure 1 Pentamidine represses proliferation of OC cells. **(A and B)** HO8910 and Caov3 cells were plated in 96-well plates and exposed to pentamidine (0, 1, 2.5, 5.0, 7.5, 15, 20 µmol/L) for 24 h in triplicates. And then cell viability was measured after adding 20 µL MTS for 1 h. Statistical analysis was performed between cells treated with 0 µmol/L pentamidine and each of the other drug concentration groups. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by one-way ANOVA. * $P < 0.05$; ** $P < 0.01$. **(C)** Colony formation assays of HO8910 and CAOV3 cells induced by pentamidine (0, 5.0, 7.5, 10 µmol/L) for 7 days. **(D and E)** The numbers of cell colonies were counted with ImageJ. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by one-way ANOVA. *** $P < 0.001$.

Pentamidine Suppresses the Cell Migration Ability of OC Cells

To evaluate whether pentamidine affected the migratory properties of OC cells, then wound healing assays and transwell migration assays were performed in HO8910 cells and Caov3 cells. In wound healing assays, pentamidine significantly inhibited cell movement and cell migration after 24 and 48 h of treatment (Figure 2A and B). The migration rate of HO8910 cells decreased from $53 \pm 1.48\%$ in the control group cells to $27.33 \pm 2.55\%$ in the pentamidine-treated cells after 48 h of treatment. The similar experiments were further performed in another OC-derived cell line, Caov3 cells. In agreement with the results generated from HO8910 cells, the data from Caov3 cells demonstrated that the migration rate reduced from $45.77 \pm 2.04\%$ in the control group to $16.87 \pm 0.23\%$ in the pentamidine treatment group after 48 h of treatment (Figure 2C and D). Meanwhile, the transwell migration assays without Matrigel showed cell migration was also suppressed by pentamidine in a dose-dependent manner (Figure 2E–G). Collectively, these results suggested that pentamidine inhibited ovarian cancer cell migration viability.

Pentamidine Upregulates PTEN Protein and Downregulates AKT Phosphorylation in OC Cells

It has been reported that phosphatase of regenerating liver-3 (PRL-3), a protein tyrosine phosphatase, downregulates PTEN expression and the activities of PRL-3 in dephosphorylating a phosphotyrosine peptide substrate in vitro are inhibited in the presence of pentamidine.^{15,16} Therefore, we wondered whether pentamidine has an effect on PTEN expression in OC cells. Firstly, Western blotting was performed to detect the influence of pentamidine on the protein expression of PTEN. HO8910 cells and Caov3 cells were exposed to increasing concentration of pentamidine for 24 h and the protein levels of PTEN were assessed. The results showed that a dose-dependent increase in the levels of PTEN (Figure 3A and B and Supplementary Figure 1). Meanwhile, quantitative Real-time PCR analysis demonstrated pentamidine did not affect the mRNA levels of PTEN, implying that up-regulation of PTEN is a posttranscriptional event (Figure 3C). Next, we turn to investigate whether pentamidine-induced upregulation of PTEN enhances its suppressive effect on PI3K.

Hence, we detected the phosphorylated and total level of AKT, and the data showed that the level of p-AKT decreased in a dose-dependent manner, while the level of total AKT did not change significantly in pentamidine-treated OC cells (Figure 3D and E and Supplementary Figure 2). These results indicated that pentamidine contributed to accumulated PTEN protein level, thus increasing inhibitory effect on PI3K/AKT signaling pathway.

Pentamidine is Involved in Maintenance of PTEN Stability via the Ubiquitin/Proteasome Pathway in HO8910 Cell Lines

To determine whether pentamidine-induced upregulation of PTEN is mediated by the ubiquitin/proteasome pathway, cells were grown in the presence or absence of pentamidine and treated with cycloheximide (CHX), a translation inhibitor. Our data showed that upon inhibition of protein synthesis by CHX, pentamidine prolonged the half-life of PTEN in HO8910 cells, suggesting pentamidine maintained the stability of PTEN (Figure 4A and B and Supplementary Figure 3). We further treated the cells with a proteasome inhibitor MG132, and pentamidine stabilized PTEN protein to a similar level as observed with MG132 (Figure 4C and D and Supplementary Figure 4). In addition, immunoprecipitation (IP)-based ubiquitination assays were performed to provide the evidence that the influence of pentamidine on PTEN ubiquitination. PTEN was immunoprecipitated with the antibody to PTEN and immunoblotted with the antibody to ubiquitin. The results showed pentamidine markedly decreased the level of PTEN ubiquitination (Figure 4E and Supplementary Figure 5). Low migrating bands corresponding to polyubiquitinated PTEN (ladders of polyubiquitinated PTEN) were detected in HO8910 cells in absence of pentamidine but were not detectable in presence of pentamidine. Taken together, these findings indicated that pentamidine maintained PTEN stability by reducing polyubiquitination of PTEN in HO8910 cells.

Effect of Pentamidine on the Expression Levels of EMT Marker Proteins in OC Cells

Having established that pentamidine inhibits ovarian cancer cell migration, we further wonder the molecular mechanisms underlying inhibitory activities. Previous

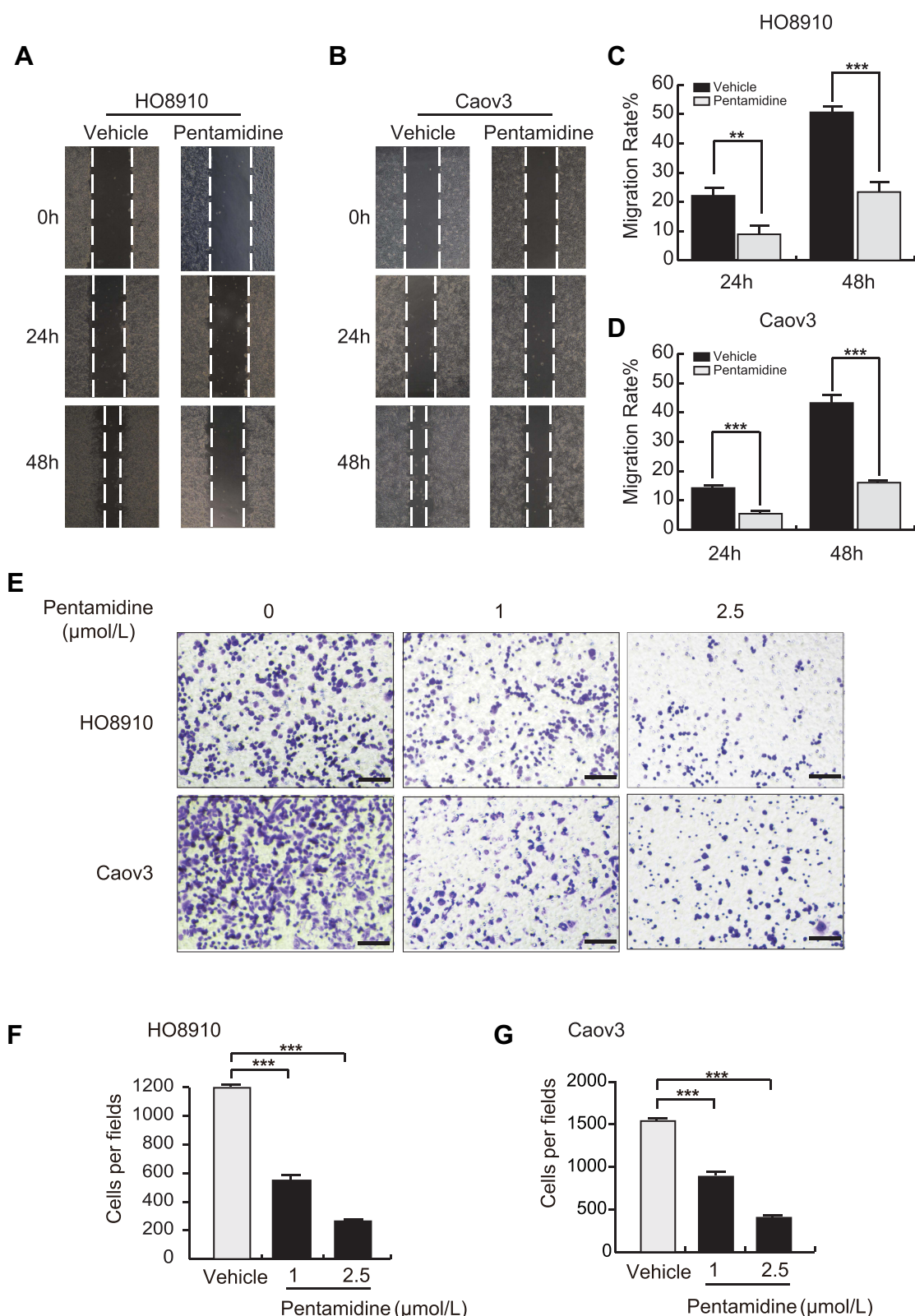


Figure 2 Pentamidine suppresses the migration of OC cells. (A and B) Wound-healing assay was performed with DMSO control and pentamidine-treated HO8910 and Caov3 cells. Representative images were obtained at 2.5 μmol/L for the indicated time. (C and D) Quantification of wounding-healing assay. Data are represented as mean ± SD from three independent experiments. Significance was determined by Student's t-test. ** $P < 0.01$; *** $P < 0.001$. (E) Transwell migration experiments of HO8910 and Caov3 cells were performed at the indicated concentrations for 6 h and representative images were photographed. Scar bars: 100 μm. (F and G) The numbers of migrating cells were counted and analyzed. Data are represented as mean ± SD from three independent experiments. Significance was determined by one-way ANOVA. *** $P < 0.001$.

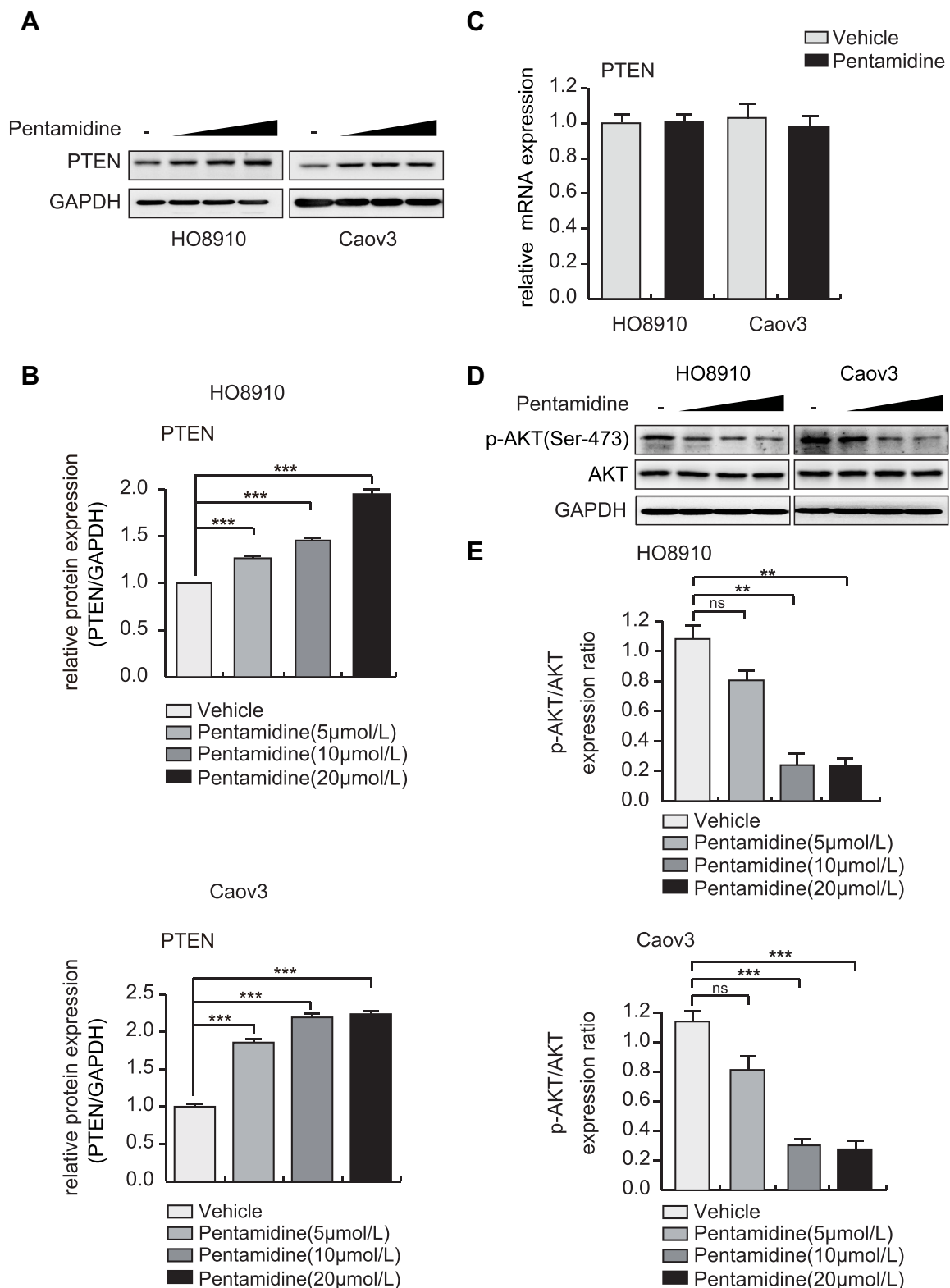


Figure 3 The effect of pentamidine on the expression of PTEN protein in OC cells. **(A)** HO8910 and Caov3 cells were treated with the increasing concentrations of pentamidine for 24 h. Cell lysates were then collected for Western blotting analysis using an anti-PTEN antibody. GAPDH was used as the loading control. **(B)** Densitometric quantification of Western blot data presented in **(A)**. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by one-way ANOVA. *** $p < 0.001$ compared to vehicle control. **(C)** HO8910 and Caov3 cells were treated with 20 μ mol/L pentamidine for 24 h and PTEN mRNA levels were determined by RT-PCR, and normalized by 18S levels. Data are represented as mean $\pm$ SD from three independent experiments. **(D)** HO8910 and Caov3 cells were treated with the indicated concentrations of pentamidine for 24 h. Protein levels of p-AKT, and AKT were analyzed by Western blotting. GAPDH was used as the loading control. **(E)** Densitometric quantification of Western blot data presented in **(D)**. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by one-way ANOVA. ** $p < 0.01$; *** $p < 0.001$.

Abbreviation: ns, no significance.

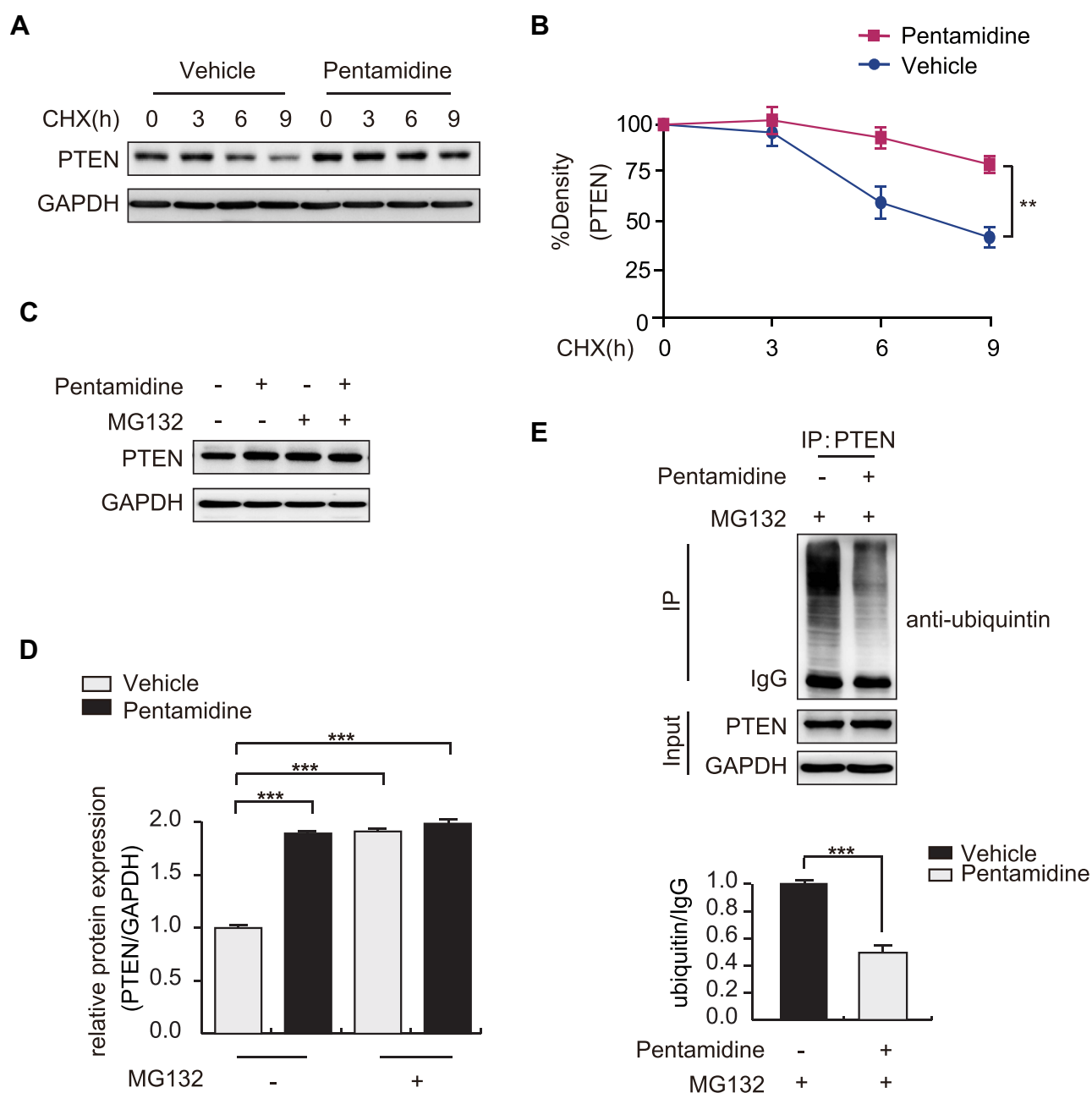


Figure 4 Effects of pentamidine on PTEN stability in human ovarian cancer HO8910 cells. **(A)** HO8910 cells were pretreated with pentamidine (20 μ mol/L) for 24 h and 80 μ g/mL cycloheximide (CHX) was added for the indicated time points, followed by Western blotting analysis. **(B)** The density of PTEN protein was expressed as a percentage of CHX-untreated control. $^{**}P<0.01$. **(C)** HO8910 cells were pretreated with pentamidine for 24 h and MG132 (20 μ mol/L) was added for 6 h, followed by Western blotting analysis. **(D)** Densitometric quantification of Western blot data presented in **(C)**. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by one-way ANOVA. $^{***}P<0.001$ compared to vehicle control. **(E)** HO8910 cells were treated with or without 20 μ mol/L pentamidine for 24 h and then MG132 (20 μ mol/L) was added for 6 h. Total lysates from cells were immunoprecipitated (IP) with anti-PTEN antibody followed by Western blotting analysis using anti-ubiquitin and anti-PTEN antibody. GAPDH was used as the loading control. Densitometric quantification of Western blot data presented in **(E)**. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by Student's *t*-test. $^{***}P<0.001$ compared to vehicle control.

studies have shown that PTEN suppresses the epithelial-mesenchymal transition (EMT) in cancer cell,^{4,17} therefore we investigated the effects of pentamidine on specific molecular associated with EMT in OC. As shown in Figure 5A–C and Supplementary Figures 6 and 7, the expression levels of mesenchymal markers N-cadherin

and snail was observed to decrease in concentration-dependent manner, whereas the expression of an epithelial marker E-cadherin increased following treatment of both HO8910 and Caov3 cell lines with pentamidine. These findings clarified that the change in EMT markers suggests pentamidine suppressed EMT, leading to loss of migration.

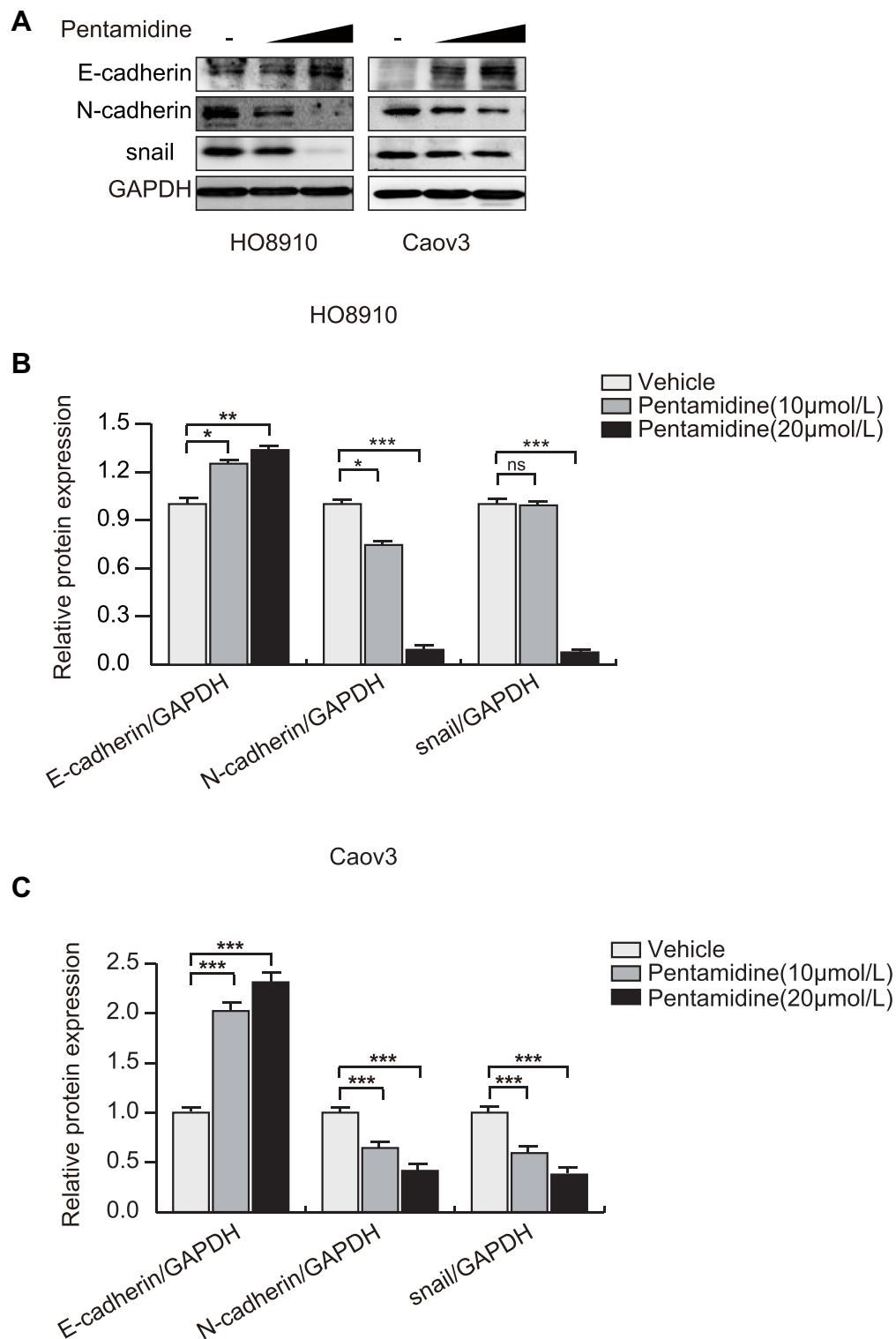


Figure 5 Pentamidine affects the expression of EMT related proteins in OC cells. **(A)** HO8910 and Caov3 cells were treated with the increasing concentrations of pentamidine for 24 h. Protein levels of E-cadherin, N-cadherin, and snail were evaluated by Western blotting. GAPDH was used as the loading control. **(B and C)** Densitometric quantification of E-cadherin, N-cadherin, and snail levels. Data are represented as mean $\pm$ SD from three independent experiments. Significance was determined by one-way ANOVA. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Abbreviation: ns, no significance compared to vehicle control.

Discussion

Pentamidine is a dicationic diarylfuran that has been studied for many decades as an antiparasitic agent.¹⁸ Recently, several studies have linked pentamidine to having anticancer effects in various cancer cells. It is reported that pentamidine can decrease HIF-1 α expression by both reducing protein stability and inhibiting protein translation in DU145 prostate and MDA-MB-231 breast cancer cell lines.¹² As well, pentamidine has been reported as a potent agent to inhibit prostate cancer progression by targeting mitochondria.⁷ In addition, in nude mice xenograft experiments, pentamidine is able to dramatically reduce the growth of WM9 human melanoma and M47 Lewis lung carcinoma tumors.^{12,13,15} These studies indicate the potential application of pentamidine in chemotherapy. Herein, we investigated the potential anticancer activity of pentamidine, in OC cell lines. Our study shows a dose-dependent suppression of pentamidine in the proliferation and migration of OC cells (Figures 1 and 2), suggesting that pentamidine acts as a potential antitumor agent that induced regression of ovarian cancer.

Indeed, even though pentamidine is used in humans to treat a number of parasitic diseases, it presents with 50% incidence of adverse effects including sudden death (at 4mg/Kg per day). In addition, some studies in literature that report the effect of pentamidine in reducing ovarian cancer or prostate cancer in mice, without side effects but do not speculate on the fact that hERG channel is not present in mice and therefore there will not be sudden death in mice but it may occur in human. Previous studies showed that the structural modifications of pentamidine differentially affected plasma membrane levels of hERG. Thus, if certain residues are avoided or “synthesized out”, the ability to inhibit trafficking can be abolished, such as pentamidine analogue (pentamidine-4, PA-4), which had mild effects on the hERG channel.¹⁹ In addition, it has been found that Dofetilide and its analogues could correct pentamidine-induced hERG trafficking defects.¹⁹ However, the effect of pentamidine and its analogues on hERG channel needs to be further evaluated, prior to clinical application of pentamidine.

PTEN, a tumor suppressor and lipid phosphatase, plays an important role in tumorigenesis by inhibiting the PI3K signaling pathway.²⁰ PTEN activity, expression and localization are regulated by post-translational modifications, including phosphorylation, ubiquitination, oxidation, acetylation and SUMOylation.^{21,22} Ubiquitination may be the

most significant post-translational modification for PTEN, which is involved in PTEN transport to different subcellular locations and regulation of its catalytic activity and stability.²³ Generally, PTEN can be regulated by both polyubiquitination and monoubiquitination.²⁴ PTEN polyubiquitination leads to its cytoplasmic retention and degradation and is therefore oncogenic in nature.²⁵ For instance, a growth factor-activated PI3K/AKT pathway suppresses MKRN1-mediated PTEN ubiquitination and degradation to accelerate tumorigenesis in cervical cancer.²⁶ On the other hand, deubiquitylase OTUS suppresses tumorigenesis and metastasis via de-polyubiquitylating and stabilizing PTEN in breast cancer.²¹ These results indicate that the tight regulation of PTEN protein stability is of great importance and more importantly, regulation in PTEN polyubiquitination levels can determine cancer outcome. In the present study, we found that the PTEN polyubiquitination level markedly decreases after pentamidine treatment compared with the control group in HO8910 cells (Figure 4E and Supplementary Figure 5), suggesting that pentamidine maybe suppress the cancer progression in OC via targeting PTEN polyubiquitination, and thus reduction in PTEN polyubiquitination level may be novel mechanism underlying pentamidine’s antitumor activity. However, further experiments are needed to elucidate the mechanism of involvement of pentamidine in regulating PTEN polyubiquitination.

The epithelial–mesenchymal transition (EMT) is the key event in distant metastasis, in which epithelial cells lose their marker proteins and acquire the mesenchymal marker proteins.¹⁷ The inactivation of PTEN may lead to the loss of apico-basal polarity and tight junctions, which are hallmarks of the EMT, which in turn promotes cell dissemination.²⁴ PTEN has been reported to be implicated in suppressing the EMT in breast cancer.⁴ In the present study, we showed pentamidine maintained PTEN stability by targeting polyubiquitination of PTEN in HO8910 cells. Moreover, that the migration of ovarian cancer HO8910 and Caov3 cells was strongly inhibited by pentamidine, as shown by the wound healing assay and transwell assay (Figure 2). In addition, we showed that pentamidine modulates the levels of EMT markers, upregulates the expression of E-cadherin, and downregulates the expression of N-cadherin and snail (Figure 5 and Supplementary Figures 6 and 7), indicating that pentamidine-induced inhibition of the migratory potential of ovarian cancer cells may be associated with the upregulation of PTEN.

Multi-drug resistance (MDR) is a common cause of the failure of chemotherapy in ovarian cancer.⁵ Patients suffering from advanced ovarian carcinoma are, in most cases, initially responsive to chemotherapy; however, they later experience a relapse of the disease because of the eventual recurrence of the tumor and emergence of drug-resistant tumor cells.²⁷ Cisplatin is an effective and widely used chemotherapeutic agent against ovarian cancer.²⁸ Resistance to platinum-based chemotherapy is a major cause of treatment failure in human ovarian cancer.⁵ Recent years, PTEN, a tumor suppressor gene, has been reported to play an important role in the reversal of MDR and become a new potential target in sensitizing cancer cells to chemotherapy.^{5,29–31} A recent study showed over-expression of PTEN is able to induce apoptosis and reverse cisplatin-resistance in ovarian cancer cell line C13K.^{5,32} Our results showed pentamidine increases PTEN protein expression in dose-dependent manner in two OC cell lines (Figure 3A and B and [Supplementary Figure 1](#)), thus providing a possibility that pentamidine treatment may sensitize OC cancer cells to chemotherapy.

Conclusions

In summary, we demonstrated for the first time that pentamidine increases PTEN expression in human ovarian cells. The up-regulation of PTEN results in inhibiting PI3K/AKT signal pathway and EMT in OC cells, thereby exerting a potent inhibitory effect on cell proliferation and migration in OC. Collectively, these results suggest that pentamidine may serve as a promising drug candidate for the treatment of OC.

Abbreviations

OC, ovarian cancer; PTEN, phosphatase and tensin homologue deleted on chromosome ten; TEPI, TGF β -regulated and epithelial cell-enriched phosphatase; MMAC1, mutated in multiple advanced cancers; ATCC, American Type Culture Collection; EMT, epithelial–mesenchymal transition; PRL-3, phosphatase of regenerating liver-3; PIP3, phosphatidylinositol (3,4,5)-trisphosphate; PIP2, phosphatidylinositol (4,5)-trisphosphate; AIDS, acquired immune deficiency syndrome; MDR, Multi-drug resistance; CHX, cycloheximide.

Funding

This work was supported by the Research Fund Liaoning Provincial Education Department (SYX202007).

Disclosure

The authors report no conflicts of interest in this work.

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